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175 mark 21 21 21	L11	(fluorescent protein or gfp) same fusion protein same (bind\$3 or interact\$3) same (fragment or domain)	660
2.00	L10	L9 same domain	187
	L9	L8 same fragment	228
	L8	L7 same fusion protein	1018
	L7	L6 same (flourescent protein or gfp)	1212
	L6	(fluorescent or gfp) same fusion same (bind\$3 or interact\$3)	3161
	L5	L1 same (fluorescent or gfp)	14
	L4	L1 same (fluorescent or gfp)	14
	L3	Li same (fluorescent or gfp)	5062
	L2	L1 and (fluorescent or gfp)	44
20 T T T	L1	protein fragment complementation assay	- 51

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=> s protein fragment complementation assay

43 PROTEIN FRAGMENT COMPLEMENTATION ASSAY

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L3 ANSWER 1 OF 8 MEDLINE on STN DUPLICATE 1

Full Text

AN 2004077559 MEDLINE

- TI Identification of an additional interaction domain in transmembrane domains 11 and 12 that supports oligomer formation in the human serotonin transporter.
- AU Just Herwig; Sitte Harald H; Schmid Johannes A; Freissmuth Michael; Kudlacek Oliver
- CS Institute of Pharmacology, University of Vienna Medical School, Wahringer Strasse 13A, A-1090 Vienna, Austria.
- SO Journal of biological chemistry, (2004 Feb 20) 279 (8) 6650-7. Journal code: 2985121R. ISSN: 0021-9258.
- CY United States
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- EM 200404
- ED Entered STN: 20040218
 Last Updated on STN: 20040501
- Entered Medline: 20040430 Na+/Cl--dependent neurotransmitter transporters form constitutive AΒ oligomers. The topological arrangement is not known, but a leucine heptad repeat in transmembrane domain (TM) 2 and a glycophorin-like motif in TM6 have been proposed to stabilize the oligomer. To determine the topology, we generated versions of the human serotonin transporter (hSERT) that carried cyan or yellow fluorescent proteins at their amino and/or carboxyl terminus. Appropriate pairs were coexpressed to measure fluorescence resonance energy transfer (FRET). Donor photobleaching FRET microscopy was employed to deduce the following arrangement: within the monomer, the amino and carboxyl termini are in close vicinity. In addition, in the oligomer, the carboxyl termini are closer to each other than the amino termini. Hence, a separate interaction domain (i.e. distinct from TM2 and TM6) must reside in the carboxyl-terminal half of hSERT. This was confirmed by expressing the amino- and carboxyl-terminal halves of hSERT. These were retained intracellularly; they also retained the coexpressed full-length transporter by forming export-deficient oligomers and, when cotransfected in all possible combinations, supported FRET. Hence, both the carboxyl and amino termini contain elements that drive oligomerization. By employing fragments comprising two neighboring TM helices, we unequivocally identified TM11/12 as a new contact site by donor photobleaching FRET and beta-lactamase protein fragment complementation assay. TM1/2 was also found to self-associate. oligomerization of hSERT involves at least two discontinuous interfaces. The currently identified interaction sites drive homophilic interactions. This is consistent with assembly of SERT oligomers in an array-like structure containing multimers of dimers.

L3 ANSWER 2 OF 8 MEDLINE on STN DUPLICATE 2

Full Text

AN 2004048932 MEDLINE

- DN PubMed ID: 14749367
- TI Regulation of apoptosis by the Ft1 protein, a new modulator of protein kinase B/Akt.
- AU Remy Ingrid; Michnick Stephen W
- CS Departement de Biochimie, Universite de Montreal, Succursale centre-ville, Montreal, Quebec H3C 3J7, Canada.
- SO Molecular and cellular biology, (2004 Feb) 24 (4) 1493-504.

 Journal code: 8109087. ISSN: 0270-7306.
- CY United States
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- EM 200403
- ED Entered STN: 20040130

Last Updated on STN: 20040313

Entered Medline: 20040312

- AΒ The serine/threonine kinase protein kinase B (PKB)/Akt plays a central role in many cellular processes, including cell growth, glucose metabolism, and apoptosis. However, the identification and validation of novel regulators or effectors is key to future advances in understanding the multiple functions of PKB. Here we report the identification of a novel PKB binding protein, called Ft1, from a cDNA library screen using a green fluorescent protein-based protein-fragment complementation assay. We show that the Ft1 protein interacts directly with PKB, enhancing the phosphorylation of both of its regulatory sites by promoting its interaction with the upstream kinase PDK1. Further, the modulation of PKB activity by Ft1 has a strong effect on the apoptosis susceptibility of T lymphocytes treated with glucocorticoids. We demonstrate that this phenomenon occurs via a PDK1/PKB/GSK3/NF-ATc signaling cascade that controls the production of the proapoptotic hormone Fas ligand. The wide distribution of Ft1 in adult tissues suggests that it could be a general regulator of PKB activity in the control of differentiation, proliferation, and apoptosis in many cell types.
- L3 ANSWER 3 OF 8 MEDLINE on STN

Full Text

- AN 2004170211 MEDLINE
- DN PubMed ID: 15064473
- TI Mapping biochemical networks with protein-fragment complementation assays.
- AU Remy Ingrid; Michnick Stephen W
- CS Departement de Biochemie, Universite de Montreal, Montreal, Quebec, Canada.
- SO Methods in molecular biology (Clifton, N.J.), (2004) 261 411-26. Ref: 26 Journal code: 9214969. ISSN: 1064-3745.
- CY United States
- DT Journal; Article; (JOURNAL ARTICLE)
 General Review; (REVIEW)
 (REVIEW, ACADEMIC)
- LA English
- FS Priority Journals
- EM 200407
- ED Entered STN: 20040406

Last Updated on STN: 20040730

Entered Medline: 20040729

AB Cellular biochemical machineries, what we call pathways, consist of

dynamically assembling and disassembling macromolecular complexes. Although our models for the organization of biochemical machines are derived largely from in vitro experiments, do they reflect their organization in intact, living cells? We have developed a general experimental strategy that addresses this question by allowing the quantitative probing of molecular interactions in intact, living cells. The experimental strategy is based on protein-fragment complementation assays (PCA), a method whereby protein interactions are coupled to refolding of enzymes from cognate fragments where reconstitution of enzyme activity acts as the detector of a protein interaction. A biochemical machine or pathway is defined by grouping interacting proteins into those that are perturbed in the same way by common factors (hormones, metabolites, enzyme inhibitors, and so on). this chapter we review some of the essential principles of PCA and provide details and protocols for applications of PCA, particularly in mammalian cells, based on three PCA reporters, dihydrofolate reductase, green fluorescent protein, and beta-lactamase.

L3 ANSWER 4 OF 8 MEDLINE on STN

DUPLICATE 3

Full Text

AN 2004114432 IN-PROCESS

- TI A cDNA library functional screening strategy based on **fluorescent** protein complementation assays to identify novel components of signaling pathways.
- AU Remy Ingrid; Michnick Stephen W
- CS Departement de Biochimie, Universite de Montreal, CP 6128, Succursale centre-ville, Montreal, Que, Canada H3C 3J7.
- SO Methods (San Diego, Calif.), (2004 Apr) 32 (4) 381-8. Journal code: 9426302. ISSN: 1046-2023.
- CY United States
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS IN-PROCESS; NONINDEXED; Priority Journals
- ED Entered STN: 20040309
 - Last Updated on STN: 20040407
- Progress towards a deeper understanding of cellular biochemical networks AB demands the development of methods to both identify and validate component proteins of these networks. Here, we describe a cDNA library screening strategy that achieves these aims, based on a protein-fragment complementation assay (PCA) using green fluorescent protein (GFP) as a reporter. The strategy combines a simple cell-based cDNA-screening approach (interactions of a "bait" protein of interest with "prey" cDNA products) with specific functional assays that use the same system and provide initial validation of the cDNA products as being biologically relevant. We applied this strategy to identify novel interacting partners of the protein kinase PKB/Akt. This method provides very general means of identifying and validating genes involved in any cellular process and is particularly designed for identifying enzyme substrates or regulatory proteins for which the enzyme specificity can only be defined by their interactions with other proteins in cells in which the proteins are normally expressed.
- ${\tt L3}$ ANSWER 5 OF 8 BIOSIS COPYRIGHT (c) 2004 The Thomson Corporation. on STN Full Text
- AN 2004:355285 BIOSIS
- DN PREV200400355536
- TI Identification of an additional interaction domain in TM11/12 that supports oligomer formation in the human serotonin transporter.
- AU Just, Herwig; Sitte, Harald; Kudlacek, Oliver kudalcek; Freissmuth,

Michael

SO Naunyn-Schmiedeberg's Archives of Pharmacology, (March 2004) Vol. 369, No. Suppl. 1, pp. R3. print.
Meeting Info.: 45th Spring Meeting of the Deutsche Gesellschaft fuer Experimentelle und Klinische Pharmakologie und Toxikologie. Mainz, Germany. March 09-11, 2004. Deutsche Gesellschaft fuer Experimentelle und Klinische Pharmakologie und Toxikologie.

ISSN: 0028-1298 (ISSN print).

DT Conference; (Meeting)

Conference; Abstract; (Meeting Abstract)

LA English

ED Entered STN: 5 Sep 2004 Last Updated on STN: 5 Sep 2004

L3 ANSWER 6 OF 8 MEDLINE on STN

Full Text

AN 2004193699 IN-PROCESS

- TI Measuring drug action in the cellular context using protein-fragment complementation assays.
- AU Yu Helen; West Mary; Keon Brigitte H; Bilter Graham K; Owens Stephen; Lamerdin Jane; Westwick John K
- CS Odyssey Thera, San Ramon, CA 94583, USA.
- SO Assay Drug Dev Technol, (2003 Dec) 1 (6) 811-22. Journal code: 101151468. ISSN: 1540-658X.
- CY United States
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS IN-PROCESS; NONINDEXED; Priority Journals
- ED Entered STN: 20040420 Last Updated on STN: 20040505
- Cellular signal transduction occurs in the context of dynamic multiprotein AΒ complexes in highly ramified pathways. These complexes in turn interact with the cytoskeleton, protein scaffolds, membranes, lipid rafts, and specific subcellular organelles, contributing to the exquisitely tight regulation of their localization and activity. However, these realities of drug target biology are not addressed by currently available drug discovery platforms. In this article, we describe the use of protein-fragment complementation assays (PCAs) to assess drugs and drug targets in the context of their native environment. The PCA process allows for the detection of protein-protein complexes following the expression of full-length mammalian genes linked in-frame to polypeptide fragments of rationally dissected reporter genes. If cellular activity causes the association of two proteins linked to complementary reporter fragments, the interaction of the proteins of interest enables refolding of the fragments, which can then generate a quantifiable signal. Using a PCA based on a yellow fluorescent protein, we demonstrate that functional (p50/p65) complexes of the heterodimeric nuclear factor-kappaB transcription factor, as well as the transcription factor subunit p65 and its modulator IkappaBalpha, can be visualized and monitored in live cells. We observed similar responses of the PCA assays to the activities of the cognate endogenous proteins, including modulation by known agonists and antagonists. A proof-of-concept high throughput screen was carried out using the p50/p65 cell line, and potent inhibitors of this pathway were identified. These assays record the dynamic activity of signaling pathways in living cells and in real time, and validate the utility of PCA as a novel approach to drug discovery.

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AN 2002301536 MEDLINE
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- DN PubMed ID: 12042868
- TI Beta-lactamase protein fragment complementation assays as in vivo and in vitro sensors of protein protein interactions.
- AU Galarneau Andre; Primeau Martin; Trudeau Louis-Eric; Michnick Stephen W
- CS Departement de Biochimie, C.P. 6128, Succursale Centre-Ville, Montreal, QC, H3C 3J7, Canada.
- SO Nature biotechnology, (2002 Jun) 20 (6) 619-22. Journal code: 9604648. ISSN: 1087-0156.
- CY United States
- DT Journal; Article; (JOURNAL ARTICLE) (VALIDATION STUDIES)
- LA English
- FS Priority Journals
- EM 200301
- ED Entered STN: 20020604

Last Updated on STN: 20030204

Entered Medline: 20030103

AB We have previously described a strategy for detecting protein protein interactions based on protein interaction assisted folding of rationally designed fragments of enzymes. We call this strategy the protein fragment complementation assay (PCA). Here we describe PCAs based on the enzyme TEM-1 beta-lactamase (EC: 3.5.2.6), which include simple colorimetric in vitro assays using the cephalosporin nitrocefin and assays in intact cells using the fluorescent substrate CCF2/AM (ref. 6). Constitutive protein protein interactions of the GCN4 leucine zippers and of apoptotic proteins Bcl2 and Bad, and the homodimerization of Smad3, were tested in an in vitro assay using cell lysates. With the same in vitro assay, we also demonstrate interactions of protein kinase PKB with substrate Bad. The in vitro assay is facile and amenable to high-throughput modes of screening with signal-to-background ratios in the range of 10:1 to 250:1, which is superior to other PCAs developed to date. Furthermore, we show that the in vitro assay can be used for quantitative analysis of a small molecule induced protein interaction, the rapamycin-induced interaction of FKBP and yeast FRB (the FKBP-rapamycin binding domain of TOR (target of rapamycin)). The assay reproduces the known dissociation constant and number of sites for this interaction. The combination of in vitro colorimetric and in vivo fluorescence assays of

beta-lactamase in mammalian cells suggests a wide variety of sensitive and high-throughput large-scale applications, including in vitro protein array analysis of protein protein or enzyme protein interactions and in vivo applications such as clonal selection for cells expressing interacting

L3 ANSWER 8 OF 8 MEDLINE on STN

DUPLICATE 5

Full Text

AN 2001431958 MEDLINE

protein partners.

- DN PubMed ID: 11479572
- TI Direct visualization of protein interactions in plant cells.
- AU Subramaniam R; Desveaux D; Spickler C; Michnick S W; Brisson N
- CS Department of Biochemistry, Universite de Montreal, Montreal, Canada H3C 3J7.
- SO Nature biotechnology, (2001 Aug) 19 (8) 769-72. Journal code: 9604648. ISSN: 1087-0156.
- CY United States
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- EM 200110
- ED Entered STN: 20011008

Last Updated on STN: 20011008 Entered Medline: 20011004

AB The protein NPR1/NIM1 is required for the induction of systemic acquired resistance (SAR) in plants and has been shown to interact with members of the TGA/OBF family of basic leucine zipper (bZIP) transcription factors. However, to date, there is no method available to monitor such interactions in plant cells. We report here an in vivo protein fragment complementation assay (PCA), based on association of reconstituted murine dihydrofolate reductase (mDHFR) with a fluorescent probe to detect protein-protein interaction in planta. We demonstrate that the interaction between Arabidopsis NPR1/NIM1 and the bZIP factor TGA2 is induced by the regulators of SAR, salicylic acid (SA), and its analog 2,6-dichloroisonicotinic acid (INA) with distinct species-specific responses. Furthermore, the induced interaction is localized predominantly in the nucleus. Protein fragment complementation assays could be of value to agricultural research by providing a system for high-throughput biochemical pathway mapping and for screening of small molecules that modulate protein interactions.

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=> s (fluorescent or gfp) and fusion and bind
          1409 (FLUORESCENT OR GFP) AND FUSION AND BIND
L4
=> s (fluorescent protein or gfp) and fusion protein and (bind or interaction)
          1896 (FLUORESCENT PROTEIN OR GFP) AND FUSION PROTEIN AND (BIND OR
               INTERACTION)
=> s 15 and fragment
L6
          148 L5 AND FRAGMENT
=> s 16 and domain
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           91 L6 AND DOMAIN
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L8 ANSWER 1 OF 78 MEDLINE on STN DUPLICATE 1 Full Text

AN 2004263910 MEDLINE

- TI A fluorescence cell biology approach to map the second integrin-binding site of talin to a 130-amino acid sequence within the rod **domain**.
- AU Tremuth Laurent; Kreis Stephanie; Melchior Chantal; Hoebeke Johan; Ronde Philippe; Plancon Sebastien; Takeda Kenneth; Kieffer Nelly
- CS Laboratoire de Biologie et Physiologie Integree (CNRS/GDRE-ITI), Universite du Luxembourg, 162A, Avenue de la Faiencerie, L-1511, Luxembourg, France.
- SO Journal of biological chemistry, (2004 May 21) 279 (21) 22258-66. Journal code: 2985121R. ISSN: 0021-9258.
- CY United States
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- EM 200406
- ED Entered STN: 20040528

Last Updated on STN: 20040701 Entered Medline: 20040630

AB The cytoskeletal protein talin, which provides a direct link between integrins and actin filaments, has been shown to contain two distinct binding sites for integrin beta subunits. Here, we report the precise delimitation and a first functional analysis of the talin rod domain integrin-binding site. Partially overlapping cDNAs covering the entire human talin gene were transiently expressed as DsRed fusion proteins in Chinese hamster ovary cells expressing alpha(IIb)beta(3), linked to green fluorescent protein (GFP). Two-color fluorescence analysis of the transfected cells, spread on fibrinogen, revealed distinct subcellular staining patterns including focal adhesion, actin filament, and granular labeling for different talin fragments. The rod domain fragment G (residues 1984-2344), devoid of any known actin- or vinculin-binding sites, colocalized with beta(3)-GFP in focal adhesions. Direct in vitro interaction of fragment G with native platelet integrin alpha(IIb)beta(3) or with the recombinant wild type, but not the Y747A mutant beta(3) cytoplasmic tail, linked to glutathione S-transferase, was demonstrated by surface plasmon resonance analysis and pull-down assays, respectively. Here, we demonstrate for the first time the in vivo relevance of this interaction by fluorescence resonance energy transfer between beta(3)-GFP and DsRed-talin fragment G. Further in vitro pull-down studies allowed us to map out the integrin-binding site within fragment G to a stretch of 130 residues (fragment J, residues 1984-2113) that also localized to focal adhesions. Finally, we show by a cell biology approach that this integrin-binding site within the talin rod domain is important for beta(3)-cytoskeletal interactions but does not participate in alpha(IIb)beta(3) activation.

L8 ANSWER 2 OF 78 MEDLINE on STN

Full Text

- AN 2004141129 MEDLINE
- DN PubMed ID: 14718533
- TI The role of the carboxyl terminus in ClC chloride channel function.
- AU Hebeisen Simon; Biela Alexander; Giese Bernd; Muller-Newen Gerhard; Hidalgo Patricia; Fahlke Christoph
- CS Institutes of Physiology, Rheinisch-Westfalische Technische Hochschule Aachen, Pauwelsstrasse 30, 52074 Aachen, Germany.
- SO Journal of biological chemistry, (2004 Mar 26) 279 (13) 13140-7. Journal code: 2985121R. ISSN: 0021-9258.
- CY United States
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- EM 200405
- ED Entered STN: 20040323

Last Updated on STN: 20040510

Entered Medline: 20040507

The human muscle chloride channel ClC-1 has a 398-amino acid carboxyl-terminal domain that resides in the cytoplasm and contains two CBS (cystathionine-beta-synthase) domains. To examine the role of this region, we studied various carboxyl-terminal truncations by heterologous expression in mammalian cells, whole-cell patch clamp recording, and confocal imaging. Channel constructs lacking parts of the distal CBS domain, CBS2, did not produce functional channels, whereas deletion of CBS1 was tolerated. ClC channels are dimeric proteins with two ion conduction pathways (protopores). In heterodimeric channels consisting of one wild type subunit and one subunit in which the carboxyl terminus was completely deleted, only the wild type protopore was functional, indicating that the carboxyl terminus supports the function of the

protopore. All carboxyl-terminal-truncated mutant channels fused to yellow fluorescent protein were translated and the majority inserted into the plasma membrane as revealed by confocal microscopy. Fusion proteins of cyan fluorescent protein linked to various fragments of the carboxyl terminus formed soluble proteins that could be redistributed to the surface membrane through binding to certain truncated channel subunits. Stable binding only occurs between carboxyl-terminal fragments of a single subunit, not between carboxyl termini of different subunits and not between carboxyl-terminal and transmembrane domains. However, an interaction with transmembrane domains can modify the binding properties of particular carboxyl-terminal proteins. Our results demonstrate that the carboxyl terminus of ClC-1 is not necessary for intracellular trafficking but is critical for channel function. Carboxyl termini fold independently and modify individual protopores of the double-barreled channel.

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L8 ANSWER 3 OF 78 MEDLINE on STN
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Full Text

- AN 2004139076 MEDLINE
- DN PubMed ID: 14699116
- TI Cytoplasmic **domain**-mediated dimerizations of toll-like receptor 4 observed by beta-lactamase enzyme **fragment** complementation.
- AU Lee Hyun-Ku; Dunzendorfer Stefan; Tobias Peter S
- CS Department of Immunology, The Scripps Research Institute, La Jolla, California 92037, USA.
- NC HL23584 (NHLBI)
- SO Journal of biological chemistry, (2004 Mar 12) 279 (11) 10564-74. Journal code: 2985121R. ISSN: 0021-9258.
- CY United States
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- EM 200405
- ED Entered STN: 20040323 Last Updated on STN: 20040520

Entered Medline: 20040519

- AB Toll-like receptors (TLRs) detect the presence of microbial challenge and initiate innate immune defensive responses. In this work we have explored the mechanism and role of TLR dimerization in signal transduction using the newly developed technique of beta-lactamase protein fragment complementation, among others. We observed that TLR4 interactions with itself, with MyD88, or with TLR2 were accurately reported by the enzyme complementation technique. That technique, as well as co-immunoprecipitation, transfection-initiated cell activation, and site-directed mutagenesis all suggest an important role for TLR intracellular domains in receptor dimerization. These findings broaden our understanding of TLR self-interactions as well as heterodimer
- L8 ANSWER 4 OF 78 MEDLINE on STN

Full Text

- AN 2004107428 MEDLINE
- DN PubMed ID: 14665628

formation.

- TI Definition of the consensus motif recognized by gamma-adaptin ear domains.
- AU Mattera Rafael; Ritter Brigitte; Sidhu Sachdev S; McPherson Peter S; Bonifacino Juan S
- CS Cell Biology and Metabolism Branch, NICHD, National Institutes of Health, Bethesda, Maryland 20892, USA.
- SO Journal of biological chemistry, (2004 Feb 27) 279 (9) 8018-28.

- Journal code: 2985121R. ISSN: 0021-9258.
- CY United States
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- OS GENBANK-AY367088
- EM 200405
- ED Entered STN: 20040305

Last Updated on STN: 20040505

Entered Medline: 20040503

- The heterotetrameric adaptor complex 1 (AP-1) and the monomeric Golgi-localized, gamma ear-containing, Arf-binding (GGA) proteins are components of clathrin coats associated with the trans-Golgi network and endosomes. The carboxyl-terminal ear domains (or gamma-adaptin ear (GAE) domains) of two gamma-adaptin subunit isoforms of AP-1 and of the GGAs are structurally similar and bind to a common set of accessory proteins. In this study, we have systematically defined a core tetrapeptide motif PsiG(P/D/E)(Psi/L/M) (where Psi is an aromatic residue), which is responsible for the interactions of accessory proteins with GAE domains. The definition of this motif has allowed us to identify novel GAE-binding partners named NECAP and aftiphilin, which also contain clathrin-binding motifs. These findings shed light on the mechanism of accessory protein recruitment to trans-Golgi network and endosomal clathrin coats.
- L8 ANSWER 5 OF 78 MEDLINE on STN

DUPLICATE 2

Full Text

- AN 2004241726 MEDLINE
- DN PubMed ID: 15141306
- TI Characterization of the yeast tricalbins: membrane-bound multi-C2-domain proteins that form complexes involved in membrane trafficking.
- AU Creutz C E; Snyder S L; Schulz T A
- CS Department of Pharmacology, University of Virginia, Charlottesville, Virginia 22908, USA.. creutz@virginia.edu
- NC GM53266 (NIGMS)
- SO Cellular and molecular life sciences: CMLS, (2004 May) 61 (10) 1208-20. Journal code: 9705402. ISSN: 1420-682X.
- CY Switzerland
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- EM 200407
- ED Entered STN: 20040514

Last Updated on STN: 20040710

Entered Medline: 20040709

AB In a survey of yeast genomic sequences encoding calcium- and phospholipid-binding C2 domains, three homologous genes were identified that encode proteins that each have three C2 domains and an apparent transmembrane domain near the N terminus. The name tricalbins is suggested for these proteins, corresponding to the open reading frames YOR086c (TCB1), YNL087w (TCB2), and YML072c (TCB3). An antiserum was raised against the C-terminal portion of tricalbin 2 and used on Western blots to demonstrate that the corresponding protein is expressed in yeast and appears as a high-molecular-weight band at 130 kDa with smaller fragments at 39 kDa and 46 kDa. A fusion protein consisting of full length tricalbin 2 fused to the green fluorescent protein was expressed in cells and found to traffic from the cell surface to intracellular vesicles near the vacuole. A two-hybrid interaction screen with the C-terminal portion of tricalbin 2 indicated that tricalbin 2 binds the C-terminal portions of tricalbins 1 and 3 suggesting that

the tricalbins may form heterodimers in vivo. Tricalbin 2 also interacted with the activation domain of the pleiotropic drug resistance transcription factor Pdrlp. Combinatorial disruptions of the tricalbin genes revealed that tcb2 single mutants or tcb1, tcb3 double mutants have an altered vacuole morphology and are hypersensitive to cycloheximide. A screen for single-copy suppressors of the cycloheximide sensitivity of tricalbin mutants yielded RSP5, which encodes a C2-domain-containing, ubiquitin-conjugating ligase essential for receptor-mediated and fluid phase endocytosis. The results suggest that the tricalbins function as multimers in membrane-trafficking events and may provide insights into the roles of multi-C2-domain proteins, such as the synaptotagmins, in other organisms.

L8 ANSWER 6 OF 78 MEDLINE on STN

Full Text

AN 2004136672 MEDLINE

DN PubMed ID: 14985083

- TI Anterograde and retrograde intracellular trafficking of fluorescent cellular prion protein.
- AU Hachiya Naomi S; Watanabe Kota; Yamada Makiko; Sakasegawa Yuji; Kaneko Kiyotoshi
- CS Department of Cortical Function Disorders, National Institute of Neuroscience, National Center of Neurology and Psychiatry, and Core Research for Evolutional Science and Technology, Japan Science and Technology Agency, Tokyo 187-8502, Japan.
- SO Biochemical and biophysical research communications, (2004 Mar 19) 315 (4) 802-7.

Journal code: 0372516. ISSN: 0006-291X.

CY United States

- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- EM 200404
- ED Entered STN: 20040320 Last Updated on STN: 20040421 Entered Medline: 20040420
- In order to investigate the microtubule-associated intracellular ABtrafficking of the NH2-terminal cellular prion protein (PrPC) fragment [Biochem. Biophys. Res. Commun. 313 (2004) 818], we performed a real-time imaging of fluorescent PrPC (GFP-PrPC) in living cells. Such GFP-PrPC exhibited an anterograde movement towards the direction of plasma membranes at a speed of 140-180 nm/s, and a retrograde movement inwardly at a speed of 1.0-1.2 microm/s. The anterograde and retrograde movements of GFP-PrPC were blocked by a kinesin family inhibitor (AMP-PNP) and a dynein family inhibitor (vanadate), respectively. Furthermore, anti-kinesin antibody (alpha-kinesin) blocked its anterograde motility, whereas anti-dynein antibody (alpha-dynein) blocked its retrograde motility. These data suggested the kinesin family-driven anterograde and the dynein-driven retrograde movements of GFP-PrPC. Mapping of the interacting domains of PrPC identified amino acid residues indispensable for interactions with kinesin family: NH2-terminal mouse (Mo) residues 53-91 and dynein: NH2-terminal Mo residues 23-33, respectively. Our findings argue that the discrete N-terminal amino acid residues are indispensable for the anterograde and retrograde intracellular movements of PrPC.

L8 ANSWER 7 OF 78 MEDLINE on STN

Full Text

AN 2004353620 MEDLINE

- TI Insulin receptor substrate of 53 kDa links postsynaptic shank to PSD-95.
- AU Soltau Michaela; Berhorster Kerstin; Kindler Stefan; Buck Fritz; Richter Dietmar; Kreienkamp Hans-Jurgen
- CS Institut fur Zellbiochemie und klinische Neurobiologie, Universitatsklinikum Hamburg-Eppendorf, Hamburg, Germany.
- SO Journal of neurochemistry, (2004 Aug) 90 (3) 659-65. Journal code: 2985190R. ISSN: 0022-3042.
- CY England: United Kingdom
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- EM 200409
- ED Entered STN: 20040717 Last Updated on STN: 20040911 Entered Medline: 20040910
- The insulin receptor substrate of 53 kDa (IRSp53) is a target of the small GTPase cdc42 which is strongly enriched in the postsynaptic density of excitatory synapses. IRSp53 interacts with the postsynaptic shank1 scaffolding molecule in a cdc42 regulated manner. The functional significance of the cdc42/IRSp53 pathway in postsynaptic sites is however, unclear. Here we identify PSD-95 as a second synaptic interaction partner of IRSp53. Interaction is mediated by a C-terminal PDZ binding motif in IRSp53 and the second PDZ domain of PSD-95. In HEK cells, overexpressed IRSp53 induces filopodia and targets PSD-95 into these processes. Immunoprecipitation and immunocytochemistry experiments demonstrate that the interaction occurs at postsynaptic sites in the brain. By virtue of its PDZ-binding and SH3 domains, IRSp53 is capable of inducing the formation of a triple complex (shank1/IRSp53/PSD-95).

L8 ANSWER 8 OF 78 MEDLINE on STN

Full Text

- AN 2004029767 MEDLINE
- DN PubMed ID: 14709720
- TI A role for the spectrin superfamily member Syne-1 and kinesin II in cytokinesis.
- AU Fan Jun; Beck Kenneth A
- CS Department of Cell Biology and Human Anatomy, School of Medicine, University of California, Davis, CA 95616, USA.
- NC GM59353-02 (NIGMS)
- SO Journal of cell science, (2004 Feb 1) 117 (Pt 4) 619-29. Journal code: 0052457. ISSN: 0021-9533.
- CY England: United Kingdom
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- EM 200409
- ED Entered STN: 20040121 Last Updated on STN: 20040930 Entered Medline: 20040929
- AB Expression of a dominant negative fragment of the spectrin family member Syne-1 causes an accumulation of binucleate cells, suggesting a role for this protein in cytokinesis. An association of this fragment with the C-terminal tail domain of the kinesin II subunit KIF3B was identified by yeast two-hybrid and co-precipitation assays, suggesting that the role of Syne-1 in cytokinesis involves an interaction with kinesin II. In support of this we found that (1) expression of KIF3B tail domain also gives rise to multinucleate cells, (2) both Syne-1 and KIF3B localize to the central spindle and midbody during cytokinesis in a detergent resistant and ATP sensitive manner and (3) Syne-1 localization is blocked by expression of KIF3B tail. Also, membrane vesicles containing syntaxin

associate with the spindle midbody with identical properties. We conclude that Syne-1 and KIF3B function together in cytokinesis by facilitating the accumulation of membrane vesicles at the spindle midbody.

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L8 ANSWER 9 OF 78 MEDLINE on STN
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Full Text

- AN 2004392329 MEDLINE
- DN PubMed ID: 15296087
- TI New chimera proteins for fluorescence correlation spectroscopy.
- AU Olah Z; Trier U; Sauer B; Schafer-Korting M; Kleuser B
- CS National Institute of Dental and Craniofacial Research, Pain and Neurosensory Mechanisms Branch, National Institutes of Health, Bethesda, USA.
- SO Die Pharmazie, (2004 Jul) 59 (7) 516-23. Journal code: 9800766. ISSN: 0031-7144.
- CY Germany: Germany, Federal Republic of
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- EM 200409
- ED Entered STN: 20040807

Last Updated on STN: 20040904

Entered Medline: 20040903

- AB A new class of chimera proteins has been developed. They are ideally suited for detection by fluorescence correlation spectroscopy (FCS), a new technology to analyze molecular interactions. The molecular structure of these chimera proteins consists of four domains: a N-terminal (His) 6-tag for affinity chromatography followed by an eight amino acid epitope for immunodetection, a polypeptide affinity domain (ADF) for target specific interaction and a C-terminal Green Fluorescent Protein (GFPuv) for reporting of interaction with the target by FCS. We designed, prepared and characterized a prototype of ADF-GFP proteins capable of specific interaction with DNA fragments bearing nuclear factor (NF)-kappaB sites. ADF NF-kappaB p50 and a non-DNA-binding deletion mutant (p35) combined with GFPuv were inserted in a procaryotic vector and expressed in E. coli. Following affinity purification the fluoroproteins p50-GFPuv and p35-GFPuv were employed in specific protein-protein and protein-DNA interaction studies. FCS analysis as well as EMSA showed that p50-GFPuv revealed a fully functional ADF. present a model for the preparation of GFP fusion proteins capable of specific interaction with proteins, lipids or nucleic acids. The rational design allows any polypeptide fragment to be incorporated into the chimeric protein. So a new series of bio-molecules with different binding specificities and assays can be developed.
- L8 ANSWER 10 OF 78 MEDLINE on STN

Full Text

- AN 2003612587 MEDLINE
- DN PubMed ID: 14576163
- Insulin-like growth factor-independent effects mediated by a C-terminal metal-binding domain of insulin-like growth factor binding protein-3.
- AU Singh Baljit; Charkowicz Dona; Mascarenhas Desmond
- CS Protigen, Inc, Sunnyvale, California 94085, USA.
- SO Journal of biological chemistry, (2004 Jan 2) 279 (1) 477-87. Journal code: 2985121R. ISSN: 0021-9258.
- CY United States
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- EM 200403

ED Entered STN: 20031230 Last Updated on STN: 20040304 Entered Medline: 20040303

Insulin-like growth factors (IGFs) play a central role in the integration AB of proliferative and survival responses of most mammalian cell types. IGF-binding protein-3 (IGFBP-3) influences IGF action directly as a carrier of IGFs but also modulates these actions indirectly via independent mechanisms involving interactions with plasma, extracellular matrix and cell surface molecules, conditional proteolysis, cellular uptake, and nuclear transport. Here we demonstrate that a short C-terminal metal-binding domain (MBD) of IGFBP-3 mediates binding to metals. MBD epitopes, sequestered in the intact molecule, are unmasked by incubation in the presence of ferrous (but not ferric or zinc) ions. An isolated 14-mer MBD peptide triggered apoptotic effects in stressed HEK293 cells as effectively as IGFBP-3. The MBD, which encompasses a nuclear localization sequence and an adjacent putative caveolin-binding sequence, mobilizes rapid cellular uptake and nuclear localization of unrelated proteins such as green fluorescent protein and streptavidinhorseradish peroxidase conjugate. Metal ions stimulate MBD-mediated cellular/nuclear uptake in vivo. Cross-linking studies showed a direct physical interaction of MBD with integrins alphav and betal, caveolin-1, and transferrin receptor. MBD-mediated protein mobilization and pro-apoptotic effects are inhibited by nystatin but not chlorpromazine, suggesting an involvement of caveolar-mediated endocytosis. However, MBD effects are inhibited by antibodies to transferrin receptor or integrins. These results are discussed with particular reference to the cell target specificity of IGFBP-3 in disease processes such as cancer and atherosclerosis.

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(FILE 'HOME' ENTERED AT 10:02:55 ON 15 OCT 2004)

FILE 'MEDLINE, BIOSIS' ENTERED AT 10:03:06 ON 15 OCT 2004 T.1 43 S PROTEIN FRAGMENT COMPLEMENTATION ASSAY L213 S L1 AND (FLUORESCENT OR GFP) L38 DUPLICATE REMOVE L2 (5 DUPLICATES REMOVED) L41409 S (FLUORESCENT OR GFP) AND FUSION AND BIND L5 1896 S (FLUORESCENT PROTEIN OR GFP) AND FUSION PROTEIN AND (BIND OR L6 148 S L5 AND FRAGMENT L7 91 S L6 AND DOMAIN L8 78 DUPLICATE REMOVE L7 (13 DUPLICATES REMOVED) => s 18 and py<1998 1 L8 AND PY<1998

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L9 ANSWER 1 OF 1 MEDLINE on STN

Full Text

AN 97075105 MEDLINE

DN PubMed ID: 8917531

TI Expression in yeast of binding regions of karyopherins alpha and beta inhibits nuclear import and cell growth.

AU Enenkel C; Schulke N; Blobel G

CS Laboratory of Cell Biology, Rockefeller University, Howard Hughes Medical Institute, New York, NY 10021, USA.

SO Proceedings of the National Academy of Sciences of the United States of America, (1996 Nov 12) 93 (23) 12986-91.

Journal code: 7505876. ISSN: 0027-8424.

CY United States

DT Journal; Article; (JOURNAL ARTICLE)

LA English

FS Priority Journals

EM 199612

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ED Entered STN: 19970128

Last Updated on STN: 20000303 Entered Medline: 19961230

AΒ Using truncated forms of recombinant yeast karyopherins alpha and beta in in vitro binding assays, we mapped the regions of karyopherin alpha that bind to karyopherin beta and the regions of karyopherin beta that interact with karyopherin alpha and with Ran-GTP. Karyopherin alpha's binding region for karyopherin beta was localized to its N-terminal domain, which contains several clusters of basic residues, whereas karyopherin beta's binding region for karyopherin alpha was localized to an internal region containing two clusters of acidic residues. Karyopherin beta's binding region for Ran-GTP overlaps with that for karyopherin alpha and comprises at least one of the two acidic clusters required for karyopherin alpha binding in addition to further downstream determinants not required for karyopherin alpha binding. Overexpression in yeast of fragments containing either karyopherin beta's binding region for alpha and Ran-GTP or karyopherin alpha's binding region for beta resulted in sequestration of most of the cytosolic karyopherin alpha or karyopherin beta, respectively, in complexes containing the truncated proteins. As these binding region-containing fragments lack other domains required for function of the corresponding protein, the overexpression of either fragment also inhibited in vivo nuclear import of a model reporter protein as well as cell growth.